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Estimation of the poroelastic parameters of cortical bone

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Abstract

Cortical bone has two systems of interconnected channels. The largest of these is the vascular porosity consisting of Haversian and Volkmann's canals, with a diameter of about $50\,\mu\text{m}$, which contains a.o. blood vessels and nerves. The smaller is the system consisting of the canaliculi and lacunae: the canaliculi are at the submicron level and house the protrusions of the osteocytes. When bone is differentially loaded, fluids within the solid matrix sustain a pressure gradient that drives a flow. It is generally assumed that the flow of extracellular fluid around osteocytes plays an important role not only in the nutrition of these cells, but also in the bone's mechanosensory system.

The interaction between the deformation of the bone matrix and the flow of fluid can be modelled using Biot's theory of poroelasticity. However, due to the inhomogeneity of the bone matrix and the scale of the porosities, it is not possible to experimentally determine all the parameters that are needed for numerical implementation. The purpose of this paper is to derive these parameters using composite modelling and experimental data from literature. A full set of constants is estimated for a linear isotropic description of cortical bone as a two-level porous medium. Bone, however, has a wide variety of mechanical and structural properties; with the theoretical relationships described in this note, poroelastic parameters can be derived for other bone types using their specific experimental data sets. © 2002 Elsevier Science Ltd. All rights reserved.

Keywords: Poroelasticity; Cortical bone; Fluid flow; Biot's theory; Mechanosensitivity; Permeability

1. Introduction

Cortical bone contains a hierarchical structure of interconnected channels (Gray's Anatomy, 1996). At the highest level, Haversian and Volkmann's canals with a diameter of about 50 µm house (a.o.) blood vessels and nerves. Canaliculi are two orders of magnitude smaller and contain the osteocytic protrusions. Being open to the vascular canals, canaliculi form a passage for nutrients and other molecules travelling to and from the osteocytes. Mechanical loading enhances the exchange of molecules by driving the free fluid in and out (Piekarski and Munro, 1977; Wang et al., 2000). Fluid flow has also been suggested to play a role in the mechanosensory system of bone (Weinbaum et al, 1991; Cowin et al., 1995; Burger et al., 1998): osteocytes thereby would be stimulated by fluid shear stress and subsequently produce signalling molecules directing the osteoclasts and osteoblasts at the bone surface. A better understanding of this mechanism is important for bonerelated clinical problems such as osteoporosis and the fixation of implants.

The relation between matrix deformation and fluid flow can be described by Biot's theory of poroelasticity (Biot, 1941, 1955). The kinematic quantities in this approach are displacement vector u_i , tracking the movement of the porous solid, and vector q_i , describing the flux of fluid mass. The field variables are total stress σ_{ij} , strain in the solid phase ε_{ij} , fluid (pore) pressure p, and specific fluid volume ζ . Strain quantities ε_{ij} and ζ are related to the displacement quantities u_i and q_i according to

$$\varepsilon_{ij} = \frac{1}{2}(u_{i,j} + u_{j,i}) \tag{1}$$

and the fluid mass balance

$$\frac{\partial \zeta}{\partial t} = q_{k,k},\tag{2}$$

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Nomenclature		σ $ au_{ m r}$	stress relaxation time
B E G K	Skempton pore pressure coefficient Young's modulus shear modulus bulk modulus	ν ζ	Poisson's ratio variation of unstrained fluid volume per unit volume of the porous material (fluid content)
k p q t u α ε φ κ μ	intrinsic (geometric) permeability pore pressure fluid flow time displacement of the solid Biot effective stress coefficient strain pore volume fraction hydraulic permeability, equal to k/μ dynamic fluid viscosity	Subscru i, j, k d u lc v f s	components of the tensors in the field equa- tions drained situation undrained situation lacuno-canalicular level vascular level fluid phase solid phase

where *t* represents time. Darcy's law relates mass flux q_i to the gradient of pore pressure *p*:

$$q_i = -\kappa p \delta_{ij},\tag{3}$$

where κ is the hydraulic permeability coefficient. κ is further defined by $\kappa = k/\mu$, with k being the intrinsic permeability and μ the fluid's dynamic viscosity. k is a function of the pore geometry, more particularly the connectedness of the porosity φ and the size and spatial arrangement of the pores.

Biot's theory further includes constitutive equations of the solid structure. Although bone is anisotropic with respect to its elastic moduli and permeability, both are considered isotropic here, because the anisotropic poroelastic bone constants have not yet been determined, and bones and osteons are almost isotropic in their transverse planes, which is where fluid flow mainly occurs. To describe the poroelastic problem, the stressstrain relations of the theory of elasticity are extended with the pressure component (Biot, 1941):

$$\sigma_{ij} + \alpha p \delta_{ij} = 2G_{\rm d} \varepsilon_{ij} + \left(\frac{2G_{\rm d} v_{\rm d}}{1 - 2v_{\rm d}}\right) \varepsilon_{kk} \delta_{ij},\tag{4}$$

where G_d and v_d are the drained shear modulus and Poisson's ratio of the solid, and α the Biot effective stress coefficient. For an ideal isotropic poroelastic material,

$$\alpha = 1 - \frac{K_{\rm d}}{K_{\rm s}} \tag{5}$$

with K_d being the drained bulk modulus of the material, and K_s that of the solid matrix (Detournay and Cheng, 1993). The constitutive relation between the variation of the fluid content ζ and stress is described by

$$2G_{\rm d}\zeta = \alpha \left(\frac{1-2v_{\rm d}}{1+v_{\rm d}}\right) \left(\sigma_{kk} + \frac{3p}{B}\right),\tag{6}$$

where B is the compressibility or Skempton pore pressure coefficient (Cowin, 1999). B is defined by (Detournay and Cheng, 1993)

$$B = \frac{\alpha K_{\rm f}}{[\alpha - \varphi(1 - \alpha)]K_{\rm f} + \varphi K_{\rm d}},\tag{7}$$

where φ is the pore volume fraction and $K_{\rm f}$ the fluid bulk modulus.

The mechanical behaviour of a fluid-saturated medium is completely described by Eqs. (1)–(7). For their application to cortical bone, parameters must be quantified that describe the linear-elastic behaviour of bone (any two of E_d , v_d , G_d , and K_d), the porosity (φ), the compressibility of the fluid phase and the solid phase (K_s and K_f), and hydraulic permeability κ . Except for φ and K_f (taken from the literature), these parameters are derived below for both levels of porosity.

2. The isotropic elastic material parameters

Cortical bone has two levels of interconnected channels. First, the vascular level porosity (diameter $\sim 50 \,\mu\text{m}$; subscript v) consisting of the space occupied by the bone fluid in the Haversian and Volkmann canals. Second, the lacuno-canalicular level porosity (diameter $< 0.5 \,\mu\text{m}$; subscript lc) containing the osteocytes and extracellular bone fluid. As it is not possible to experimentally determine the elastic material constants of the solid matrix at these levels, a theoretical approach is presented, based on experimental data taken from literature. For reasons given earlier, the approach is limited to isotropic conditions.

Cowin and Sadegh (1991) constructed a set of effective isotropic elastic constants for human cortical bone, based on experimental orthotropic data from Ashman et al. (1984). It is assumed here that the vascular pores were no longer saturated, so the fluid pressure is zero. Consequently, the effective elastic Table 1 Summary of the isotropic elastic constants for cortical bone on two levels of porosity^a

Property (units, if any)	Vascular level	Lacuno- canalicular level
φ , porosity	0.04	0.05
$K_{\rm f}$ (GPa), bulk modulus of fluid	2.3	2.3
$E_{\rm d}$ (GPa), drained Young's modulus	14.58	15.75
v _d , drained Poisson's ratio	0.325	0.325
$G_{\rm d}$ (GPa), drained shear modulus	5.50	5.94
$K_{\rm d}$ (GPa), drained bulk modulus	13.92	14.99
$E_{\rm s}$ (GPa), solid Young's modulus	15.85	17.51
v _s , solid Poisson's ratio	0.333	0.335
$G_{\rm s}$ (GPa), solid shear modulus	5.94	6.56
K _s (Gpa), solid bulk modulus	15.82	17.66
$E_{\rm u}$ (GPa), undrained Young's modulus	14.65	15.85
$v_{\rm u}$, undrained Poisson's ratio	0.332	0.333
$G_{\rm u}$ (GPa), undrained shear modulus	5.50	5.94
$K_{\rm u}$ (GPa), undrained bulk modulus	14.56	15.82
α , effective stress coefficient	0.120	0.151
<i>B</i> , Skempton pore pressure coefficient	0.367	0.344

^a The values in italic were taken from Cowin (1999). The other elastic constants have been calculated from Eqs. (8)–(12). The expressions for α and *B* can be found in Detournay and Cheng (1993), p. 121 and 118, respectively.

constants are the drained elastic moduli at the vascular level: $K_{d,v} = 13.92 \text{ GPa}$, $G_{d,v} = 5.50 \text{ GPa}$, $E_{d,v} = 14.58 \text{ GPa}$, and $v_{d,v} = 0.325$ (Table 1). The vascular porosity $\varphi_v = 0.04$ (Zhang et al., 1998).

The theory of composite materials (Christensen, 1979) provides equations relating effective elastic constants of porous materials to the size and distribution of the pores and the elastic properties of the matrix material. The following relationships between the drained moduli ($K_{d,v}$ and $G_{d,v}$) and the solid phase moduli ($K_{s,v}$, $G_{s,v}$) are applications of equations in Christensen (1979) for spherical voids:

$$K_{\rm d,v} = K_{\rm s,v} - \frac{K_{\rm s,v}\varphi}{1 - K_{\rm s,v}/(K_{\rm s,v} + (4/3)G_{\rm s,v})}$$
(8)

and

$$\frac{G_{\rm d,v}}{G_{\rm s,v}} = 1 - \frac{15(1 - v_{\rm s,v})\varphi}{7 - 5v_{\rm s,v}}.$$
(9)

Further, for all isotropic linear elastic materials,

$$\frac{K_{\rm s,v}}{G_{\rm s,v}} = \frac{2(1+v_{\rm s,v})}{3(1-2v_{\rm s,v})}.$$
(10)

Using (8)–(10), the unknowns $K_{s,v}$, $G_{s,v}$ and $v_{s,v}$, as well as $E_{s,v}$ are determined: $K_{s,v} = 15.82$ GPa; $G_{s,v} = 5.94$ GPa; $v_{s,v} = 0.333$; and $E_{s,v} = 15.85$ GPa (Table 1).

The elastic constants of the porous bone material at the lacuno-canalicular level can be determined similarly. As the experiments by Ashman et al. (1984) were performed with ultrasound (2.25 MHz), and the relaxation time of the lacuno-canalicular fluid pressure is of the order of 0.1–3.0 s (Pienkowski and Pollack, 1983; Otter et al., 1992), the porous structure is now considered *undrained*. Consequently, the undrained elastic moduli for the porous medium at the lacunocanalicular level equals the elastic moduli for the solid material at the vascular level ($K_{u,lc} = K_{s,v}$; $G_{u,lc} = G_{s,v}$; $v_{u,lc} = v_{s,v}$; $E_{u,lc} = E_{s,v}$). Detournay and Cheng (1993) give the following relationships between undrained, drained and solid elastic moduli:

$$K_{\rm u} = K_{\rm d} \left[1 + \frac{(1 - K_{\rm d}/K_{\rm s})^2}{K_{\rm d}/K_{\rm s} (1 - K_{\rm d}/K_{\rm s}) + \varphi(K_{\rm d}/K_{\rm f} - K_{\rm d}/K_{\rm s})} \right]$$
(11)

and

$$v_{\rm u} = \frac{3K_{\rm u} - 2G_{\rm d}}{2(3K_{\rm u} + G_{\rm d})}.$$
(12)

Combining (11) with (8), and (12) with (9) and (10), two expressions for K_u are obtained with two unknowns, K_s and v_s . With $\varphi_{lc} = 0.05$ (Zhang et al., 1998), the elastic moduli for the solid material at the lacuno-canalicular level are: $K_{s,lc} = 17.66$ GPa; $G_{s,lc} = 6.56$ GPa; $v_{s,lc} = 0.335$; and $E_{s,lc} = 17.51$ GPa. Using (8)–(10), the drained elastic moduli become: $K_{d,lc} = 14.99$ GPa; $G_{d,lc} = 5.94$ GPa; $v_{d,lc} = 0.325$; and $E_{d,lc} = 15.75$ GPa. Note that the solid moduli are higher at levels of lower porosity: $K_{s,bone} < K_{s,v} < K_{s,lc}$.

Many other parameters can be used to describe the poroelastic problem, such as the effective stress coefficient α (Nur and Byerlee, 1971). Also, the Skempton pore pressure coefficient *B* and the undrained elastic material properties can be calculated if a value for the bulk modulus of the contents of the porosities is assumed (Detournay and Cheng, 1993). Table 1 gives a summary of these.

3. The hydraulic permeability of the lacuno-canalicular network

In porous media, fluid flow depends on the pressure gradient, fluid viscosity μ , and the structure of the porosity, expressed as intrinsic permeability k. k depends on the number, orientation and size of the canaliculi, as well as on the amount of filling by osteocytes and their processes. These parameters are



Fig. 1. Relaxation curve of the streaming potentials after step loading recorded in a dog's femur in vivo. Adapted from Otter et al. (1992).

very difficult to determine, leading to estimations of permeability k ranging over several orders of magnitude (Wang et al., 1999). Therefore, a more direct approach is chosen to estimate the hydraulic permeability κ , which includes both the intrinsic (geometrical) permeability k and the fluid viscosity μ ($\kappa = k/\mu$).

In a porous structure loaded by a step-function, fluid is pressurised at t = 0, and then relaxes due to drainage; in osteonic bone, extracellular fluid drains into the Haversian canals (Starckebaum et al., 1979; Iannacone et al., 1979). The relaxation curve is basically a function of the osteonic dimensions; the material properties of the solid bone matrix and the extracellular fluid; the porosity of the bone matrix; and the hydraulic permeability of the system. Except for the hydraulic permeability, all these parameters can be estimated using data from the literature or Section 2. Hydraulic permeability then can be estimated with the use of a finite element model of an osteon, by relating the calculated step response with an experimentally determined relaxation curve. For this study, we compare with the relaxation curve of streaming potentials recorded in vivo in a dog femur (Otter et al., 1992) (Fig. 1).

An axisymmetric finite element model of an osteon was built with an inner and outer radii of 12.5 and 80 µm, respectively. These are characteristic dimensions of an osteon in a dog's femur (Jowsey, 1966). The finite element code DIANA (TNO, The Netherlands) requires the drained Young's modulus ($E_{d,lc} = 15.75$ GPa) and Poisson's ratio ($v_{d,lc} = 0.325$); the lacuno-canalicular bone porosity ($\varphi_{lc} = 0.05$; Zhang et al., 1998); the bulk moduli of the solid matrix ($K_{s,lc} = 17.66$ GPa) and the extracellular fluid ($K_f = 2.3$ GPa; Anderson, 1967); and the hydraulic permeability, the value of which is to be determined.

Based on experiments by Starckebaum et al. (1979), Iannacone et al. (1979) and Otter et al. (1994), it is assumed that every osteon is a draining system on its own. Consequently, the outer boundary is considered



Fig. 2. (a) Relaxation of the normalised pressure difference between the inner and outer osteon border for different values of the hydraulic permeability κ . The experimental relaxation curve by Otter et al. runs between the curves for $\kappa = 10^{-6}$ and $10^{-7} \text{ mm}^4/\text{N s}$. (b) Further refinement results in an estimation of the hydraulic permeability $\kappa = 2.2 \times 10^{-7} \text{ mm}^4/\text{N s}$.

impermeable. The cylinder is further considered to be infinitely long, so only the surface of the Haversian canal is open for drainage. The reference pressure within the channel was set to zero (Cowin, 1999), and an axial compressive step load was applied resulting in a deformation of 1000 µstrain. The model was analysed with $\kappa = 10^{-6} \text{ mm}^4/\text{N s}$, $\kappa = 10^{-7} \text{ mm}^4/\text{N s}$, and $\kappa =$ $10^{-8} \text{ mm}^4/\text{N s}$. The estimation was subsequently refined in analyses with intermediate values. As drainage of bone fluid is considered to occur exclusively through the lacuno-canalicular network into the Haversian canal, the relaxation of the streaming potential determined by Otter et al. is proportional to the relaxation of the pressure gradient over the osteonic wall.

Fig. 2 shows the normalised pressure drop, with unity at t = 0.1 s. Note that the experimentally determined relaxation curve by Otter et al. has an offset of ~7 μ V



Fig. 3. (a) The experimental and numerical curves on a logarithmic scale are well approximated by a normalised negative e-power with a time constant $\tau_r = 0.85 \,\text{s.}$ (b) The slope of the logarithmic curves shows, however, that both the experimental and the numerical curve have time constants increasing from approximately $\tau_r = 0.60$ at $t = 0.1 \,\text{s}$ to $\tau_r = 0.95 \,\text{s}$ after $t = 2.0 \,\text{s.}$

(Fig. 1); as no rheological reason exists for this threshold, this value is considered as a zeroing error, and subtracted from the normalised curve. Fig. 2a shows the experimental curve between the numerical curves for $\kappa = 10^{-6}$ and 10^{-7} mm⁴/N s. Further refinement results in an estimation of $\kappa = 2.2 \times 10^{-7} \text{ mm}^4/\text{Ns}$ (Fig. 2b). On a logarithmic (ln) scale, relaxation is described by an almost straight line, which is well approximated by a normalised negative e-power with time constant $\tau_{\rm r} = 0.85$ s (Fig. 3a). However, the slope of the logarithmic relaxation function shows that there is not one single time constant, but—at least within the time frame up to 2 s—a shifting time constant (Fig. 3b). Kufahl and Saha (1990) explained this phenomenon by assuming that the deeper parts of the osteon relax more slowly than the superficial parts of the bone, which we found in our model as well (Fig. 4). The question of biphasic relaxation curve in pore fluid pressure is also measured and discussed in the work of Qin et al. (2001, 2002).



Fig. 4. Relaxation curves at different sites along the osteonic radial during the first second after loading. Note that the deeper sites start to relax later than the more superficial bone layers. Also note that most of the fluid flow occurs at the bone surface near the Haversian canal. Depths are in μm .

4. Discussion

The parameters for a double porosity isotropic Biot model of cortical bone were estimated using composite modelling and experimental data from literature. The theoretical part is based on the theory of poroelasticity (Biot, 1941; Detournay and Cheng, 1993), and contains two assumptions that have to be considered.

First, the two porosity levels in cortical bone are considered to act independently, because the pressure relaxation times differ by four orders of magnitude. This means that it is reasonable to assume that the bone matrix with the lacuno-canalicular network can be considered solid for the short time scale behaviour of the vascular porosity, and the pressure in the vascular porosity does not interfere with the fluid flow within the lacuno-canalicular network. The relaxation of the streaming potentials measured by Otter et al. (1992) is assumed to originate from the lacuno-canalicular network only, as the pressure within the vascular porosities must be completely relaxed after about 100 μ s.

The second assumption is isotropy. For the elasticity parameters, this is justified even for lamellar bone (Cowin and Sadegh, 1991), but permeability must differ considerably between longitudinal and radial directions; transverse and longitudinal sections of lamellar cortical bone give the impression that canaliculi mainly run radially towards the Haversian canals (Gray's Anatomy, 1995). With the axisymmetric approach in the finite element analysis, the estimated permeability thus is in the transverse plane of the osteon.

The value for the hydraulic permeability estimated in this study ($\kappa = 2.2 \times 10^{-7} \text{ mm}^4/\text{N s}$) is two orders of magnitude smaller than those used in studies by Zhang et al. (1998) and Wang et al. (1999). These authors based

their estimation on geometrical parameters of the porosity, the values of which are in fact highly uncertain. One physical explanation for this discrepancy could be that the degree of filling of the canaliculi by the osteocytic processes was underestimated, or in other words, the fluid layer between the cellular processes and the bone matrix is less than that assumed by these authors (some 50 nm). This would fit with the findings by Petrov (2000) that the effective porosity radius in bone is of the order of 10 nm. Other possible explanations are that the viscosity or the number of canaliculi is higher. On the other hand, the present study also has debatable input values. Lacuno-canalicular porosity, for example, was assumed to be 5%, but has also been reported to be as low as 0.023 (in man; Frost, 1960) and 0.042 (in dogs; Morris et al., 1982). The poroelastic bone parameters and the hydraulic permeability, however, are not very sensitive to changes in the lacuno-canalicular porosity, that is, their values remain of the same order of magnitude (own unreported data; Zhang et al., 1998). The same holds for osteonic dimensions and the elastic moduli of the bone tissue. It should be noted, however, that the porosity of 5% refers to the whole lacunocanalicular space, and includes both the cells and the freely flowing fluid. Cells and fluid are combined here because of their comparable bulk modulus, as opposed to the bulk modulus of the mineralised matrix. The absolute fluid flow thus will be higher than calculated with the model presented, because a part of the porosity contents does not flow. The estimated hydraulic permeability, however, does not change because it has been derived directly from the relaxation times, not from the geometrical dimensions of the canaliculi and osteocytic protrusions.

It is interesting to note that the essentials of the relaxation curve recorded in vivo by Otter et al. can be approximated with a simple finite element model of fluid flow in an osteon under axial step compression. Especially, the observation that the relaxation curve is described by a shifting time constant due to the later response of the deeper osteon layers, strongly suggests that the streaming potentials measured by Otter et al. are due to canalicular fluid flow *only*, and do not require a pathway through or an exchange with the bone's microporosity, as suggested by others (Salzstein et al., 1987; Pollack and Petrov, 2000; Petrov and Pollack, 2000). This hypothesis is consistent with observations by Otter et al. (1994) that the bone fluid mainly drains into the Haversian canals.

Finally, it must be considered that cortical bone may show a wide variety of elastic moduli, permeabilities and osteonal dimensions for functional reasons. It is conceivable, for example, that the human skull has different values as compared to the rest of the skeleton. In this context, it is also interesting to consider that osteonic dimensions differ with species (Jowsey, 1966; Albu et al., 1990), but also differ within the same animal (Skedros et al., 1997). What this means for the mechanosensitivity of their bone tissue is an interesting area of research.

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