Yeast Miniprep

1. Grow cells in 3ml selective media to an OD$_{600}$ > 1.
   Overnight - use all 3ml
   Two days - use 1.5ml
2. Pellet cells in a 1.5ml microfuge tube.
3. Resuspend by vortexing in 200ul of SCE/Zymolyase/2ME. Incubate at 37C 30 - 60 minutes.
4. Add 400ul 0.2N NaOH/1% SDS (made fresh). Invert to mix. Incubate on ice 5 minutes.
5. Add 300ul cold 3M K/5M OAc. Invert to mix. Incubate on ice 5 minutes.
6. Spin 2 minutes at top speed in a microcentrifuge. Pour supernatent into a fresh tube. Repeat spin. Remove 500ul to a fresh tube.
7. Add 300ul isopropanol. Vortex and let stand for 5 minutes at room temperature.
8. Spin at top speed for 5 minutes at room temperature. Pour off supernatent and wash pellet with 0.5ml 70% ethanol. Repeat spin.
9. Dry pellet and resuspend in 25ul TE.
10. Transform into E. coli. Use 1ul for electrocompetent cells or 10ul for chemically competent cells.

SCE Solution

1M sorbitol 100ml
18.2g
0.1M sodium citrate pH 7.6 2.94g
0.06M EDTA 2.23g

SCE/Zymolyase/2ME Solution

5ml SCE
60ul 10mg/ml Zymolyase (in 1M Sorbitol)
10ul 2-mercaptoethanol

NaOH/SDS Solution (made fresh)

100ul 10N NaOH
500ul 10% SDS
4.4ml dH$_2$O

3M K/5M OAc

60ml 5M potassium acetate
11.5ml glacial acetic acid
28.5ml dH$_2$O